

# LIFE OrgBalt progress and what`s next – overall view

#### **3<sup>rd</sup> Steering group meeting** February 4th, 2021, Teams

**Kaido Soosaar** Tartu University

LIFE OrgBalt, LIFE18 CCM/LV/001158

EU LIFE Programme project "Demonstration of climate change mitigation potential of nutrients rich organic soils in Baltic States and Finland"







Latvia University of Life Sciences and Technologies











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#### Content

#### WG measurements activities

- How we work (subgroups)
- Measurements protocols
- Sites establishment
- Measurements of GHG, biomass, environmental parameters
- Raw data storage





### Formation of subgroup

Work package	Subgroup leader	Participant	
1) Site preparations	Jyrki Jauhiainen	FI: Jyrki J, EE: Kaido S	LV: Andis L, Mārtiņš V LI: Egidijus V.
2) Heterotrophic CO <sub>2</sub> flux monitoring	Päivi Mäkiranta	FI: Päivi M EE: Ain K, Kaido S	LV: Andis L, Mārtiņš V LI: Dovilė Č, Egidijus V.
3) Transparent chamber measurements (CO <sub>2</sub> )	Kaido Soosaar	FI: Sanna S, Saara L EE: Ain K	LV: Andis L, Mārtiņš V LI: Egidijus V
4) Static dark chamber monitoring (incl. CH <sub>4</sub> & N <sub>2</sub> O)	Ain Kull	FI: Päivi M EE: Thomas S	LV: Andis L, Mārtiņš V LI: Dovilė Č, Egidijus V.
5) Meteorological parameters	Thomas Schindler	Fi: Päivi M EE: Kaido S	LV: Mārtiņš V LI: Dovilė Č
6) Water & soil, litter sampling	Mārtiņš Vanags-Duka	Fl: Timo P EE: Ain Kull	LV: Aldis B LI: Kęstutis A.
7) Litter production and decomposition belowground	Raija Laiho	FI: Tuula L EE: Ivika O	LV: Andis L, Mārtiņš V LI: Dovilė Č
8) Biomass production aboveground	Andis Lazdiņš	FI: Timo P EE: Ivika O	LV: Mārtiņš V LI: Olgirda B., Vaiva K
9) Data management (codes and storage)	Aldis Butlers	FI: Jyrki J EE: Kaido S	LV: Mārtiņš V LI: Vaiva K
10) Microbiology (New – to be formed ASAP)	Jyrki Jauhiainen	Fl: Hannu F, Krista P EE: Mikk E	LV: LI:
11) FTIR (New– to be formed ASAP)	Jyrki Jauhiainen	FI: Jyrki J FF: Ain K	LV: Aldis B LI: Dovile C





## Field protocols

1st step: to harmonize the field measurements technique and 2 field protocols have been developed:

- 1) Biomass and litter decomposition measurement protocols
- 2) Flux and Environmental data protocols

Supportive activities:

- Field Calibration seminar in Tartu 29-30.6.2020
- Workshop about harmonization of field measurements in Kaunas 25-26.08.20
- Many meetings and emails

Biomass and litter decomposition measurement protocols draft 1		
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Schedule of biomass and biomass production & decomposition studies	2	
Vegetation biomass (Kestutis A. et al.)	2	
Tree stand biomass and biomass production (incl. coarse root)	2	
Ground vegetation biomass	2	
Ground vegetation biomass production	3	
Annual herbaceous ground vegetation biomass production	3	
Perennial ground vegetation biomass production	3	
Moss biomass production	3	
Ground vegetation coverage (optional)	5	
Litter production and decomposition (Raija L. et al.)	5	
Aboveground litter production	6	
Annual tree mortality	6	
Production of aboveground tree litter	6	
Production of aboveground litter of vascular ground vegetation	6	
Production of moss litter	7	
Aboveground litter decomposition	7	
Belowground biomass, biomass production, litter production and litter decomposition	.9	
Fine root biomass	9	
Fine-root production	10	

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Soil GHG balance monitoring method harmonization in L	TPP	
OrgBalt	u.F.	
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#### Establishment of sites

By the end of January 2021 all the study sites have been selected and prepared for the field measurements.





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#### Ecosystem C and N balance



Figure 1. CO<sub>2</sub>, CH<sub>4</sub>, and N<sub>2</sub>O fluxes and mass transfer components (arrows indicate flux/transfer direction) contributing to soil C-stock changes in a forest ecosystem on drained organic soil (as in IPCC, 2014), modified from Jauhiainen et al. (2019).



#### Basic scheme of the study site





In addition, periodically soil and water samples are taken for the chemical analyzes!



#### Gaseous fluxes - $CH_4 \& N_2O$ (& winter $CO_2$ ) fluxes Monitoring with closed static chamber method: 24 months (2021-2022)



#### Measurements:

- In Finland: May 2020
- In Estonia and Latvia: Jan 2021
- In Lithuania: Feb 2021
   Frequency (minimum)
- Winter: once per month
- Vegetation period: once every three weeks



### Gaseous fluxes – heterotrophic CO<sub>2</sub> flux

Monitoring soil heterotrophic CO<sub>2</sub> fluxes on forest floor: 2 vegetation periods (2021-2022)







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Monitoring soil heterotrophic  $CO_2$  fluxes on forest floor: 2 vegetation periods (2021-2022)



Dynamic closed chamber method (EGM5(4)/Licor analyzer) Measurements :

- Finland: spring 2020
- In Estonia, Latvia and Lithuania: spring 2021 Frequency (minimum):
- during the vegetation period once every three weeks





# Gaseous fluxes - NEE from non-forested sites

#### NEE measurements from non-forested sites with transparent chamber: 2 years



#### Dynamic closed chamber method (EGM5(4) analyzer)



Measurements :

 In Estonia, Latvia and Lithuania: spring 2021

Frequency (minimum):

 during the vegetation period once every three weeks



#### C/N input - Annual litter production in forest ecosystems

Annual litter production will be **calculated as the sum of aboveground litter of tree stand, vascular ground vegetation and mosses and belowground litter of roots and rhizomes.** 

**ABOVEGROUND LITTER OF TREE STAND** 

• Fine woody litter (twigs, branches)



Stand foliar and cone litter





#### C/N input - annual litter production

Annual litter production will be calculated as the sum of **aboveground litter** of tree stand, vascular ground vegetation and **mosses** and belowground litter of roots and rhizomes.

Moss biomass production, which is assumed to equal litter production, is measured with square shaped nets (each about 20 cm x 20 cm). Will be placed on patches of three most common moss species or moss patch types (five nets per species/patch type) at each site in the autumn of the first GHG monitoring year.





### C/N input - annual litter production

Annual litter production will be calculated as the sum of **aboveground litter of** tree stand, **vascular ground vegetation** and mosses and **belowground litter of roots and rhizomes**.

- Ground vegetation biomass is harvested from six 30 x 30 cm (area = 900 cm2) areas per site, two at each GHG measurement transect. Biomass samples will be harvested during the maximum biomass in July-August.
- Fine-root production is estimated using the ingrowth-core method for peat soils. The amount of ingrown roots represents fine-root production over the incubation period, which will be generalized into annual production.





### Sampling protocols

The avoid losses and harmonize the results several sampling protocols have been developed.

Site ID: EEC1101			Date and ti	ime:
Person name(s):				
Weather conditions	2			
description				
Air temp on site				
Soil Temp:	D	Α	В	С
10 cm				
20 cm				
30 cm				
40 cm				
Soil moisture (5cm)				
WL manual (cm):				
Tube insight				
Tube outsight				
WL auto (B)				
Snow depth (cm)				
Chamber volume change				
(filled with snow or water				
(chamber volume and				
minus volume filled with				
snow/water)				
Vegetation description				
(harvesting):				
Equipm. Used (deviation fro	om standard i	nstrumentatio	on):	

**Remarks comments** (e.g. disturbances caused by animals/trespassers/weather; deviations from standard monitoring plan, use of extensions):

A STATE	
3.	
SIL AVA	



Latvian State Forest Res	earch Institute "SILAVA"
Fore	st environment laboratory
Riga stre	et 111, Salaspils, LV - 2169

#### VL\_55 Soils sampling protocol

	Sample code : OrgBalt		
a 11 1	(Specified in testing application)		
Sampling plan	Refference to sampling plan:		
	Person responsible:		
~ · · · · ·		& analyses	
		z kit	
Latvian S	State Forest Research Institute "SILAVA"	nsulation 🗆	Other:
	Forest environment laboratory	·	
	Riga street 111, Salaspils, LV - 2169		

#### VL\_51 Water sampling protocol

	Sample code : Or	gBalt			
Sampling plan	Reference to same	aling plan:			
	Demon company	ai ai		š	Date and tin
	Ferson responsion	e.		1 40 40 50	
Sampling method		M07 Periodic water sampling &	analyses	7-40, 40-30	
Identification of equipment		Water sampling kit			
Transportation conditions	Thermal box $\Box$	Box without thermal insulation $\Box$	Other:		
Atypical environmental conditions, which may affect the sample	No 🗆	Observation:			
Deviations*	No 🗆	Yes:			
Sampler	Name Surname:				
Deviations, additions or exceptions to th	e sampling method a	nd sampling plan			

ple iden	tification	S	ample iden	tification
ibplot	Date and time	Site ID	Subplot	Date and time
	ple ider bplot	ple identification           bplot         Date and time	ple identification     S       bplot     Date and time     Site ID	pleidentification         Sample identification           bplot         Date and time         Site ID         Subplot

Fails: VL_51_udens_nem_prot.docx Versija: 02	Water sampling protocol

5	Date and time
-40, 40-50	

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#### Raw data storage

- Raw data will be stored in LIFE OrgBalt SharePoint (accessible for all the partners)
- All the activities (soil sampling, water sampling, etc.) will be coded, so we know exactly from where and what has been measured.
- However, the final format of the data storage is under process
- To keep everybody updated, our working group helds monthly meetings where we cover all the needed aspects.

CISER DrgBalt Private group		
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Home		
Conversations	Documents > WG Measurements	
Documents	$\square$ Name $\vee$	Ν
Shared with us	Data storage	Д
Notebook	Field protocols from Silava	Д
Pages	OrgBalt measurements protocol	le
Site contents	OrgBalt sites	le
Recycle bin	WG Measurements meetings	le
Edit	Auxiliary data recorded during each site visi	Ji
	LIFE_Field _protocol_template.docx	k





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